

# BIOMEDICAL APPLICATIONS OF MICROPROBE ANALYSIS

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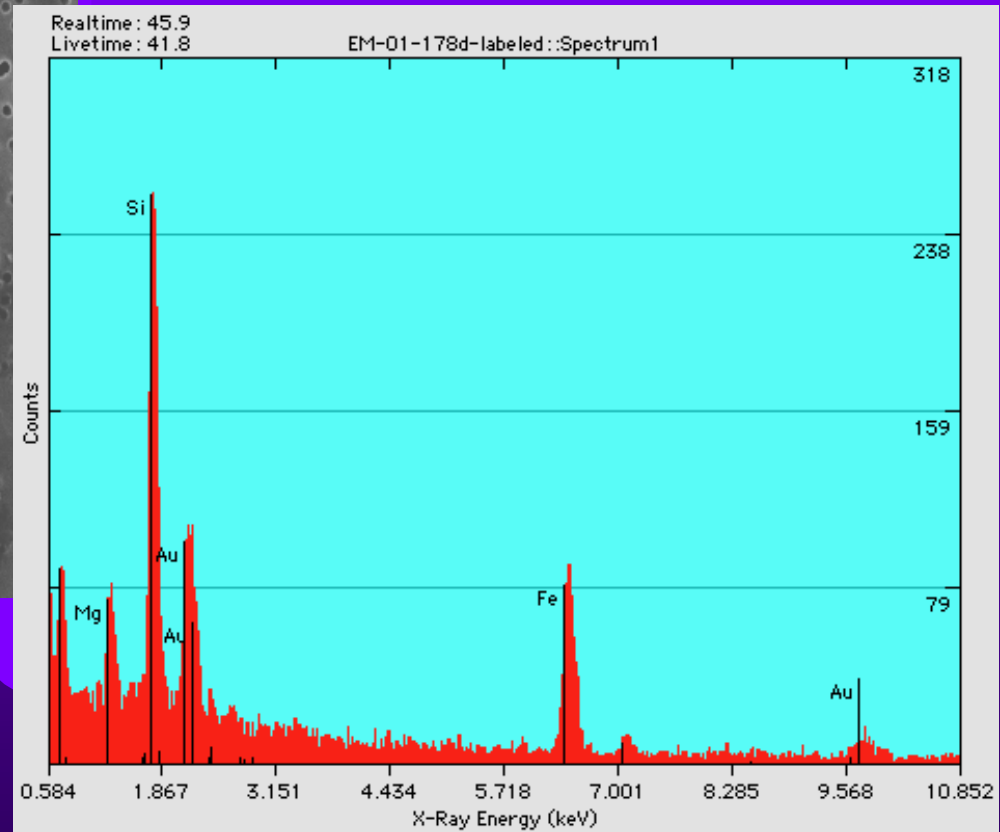
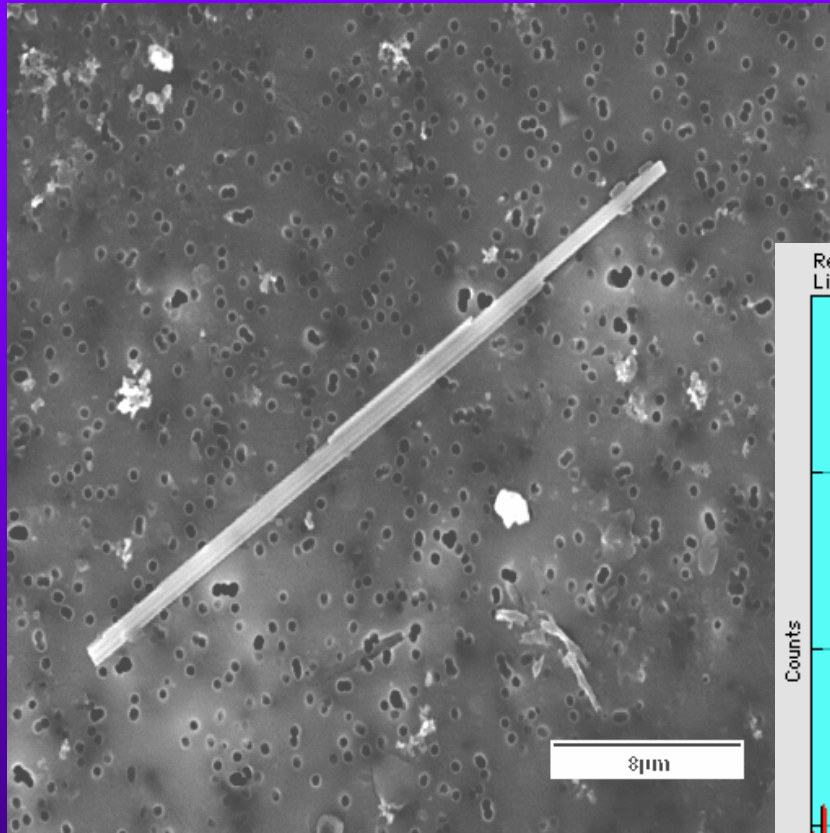
\* Use of the Advanced Photon Source was supported by the U.S. Department of Energy, Basic Energy Sciences, Office of Energy Research, under Contract No. W-31-109-Eng-38



# Microanalysis as a Diagnostic Standard of Care

- Unexplained Particulates, Inclusions, Granulomas
- **Mineral Pneumoconioses (Asbestosis)**
- **Renal Stone Disease**
- Reactions to Element-Based Anti-arrhythmics, Chemotherapeutics, Contrast Agents, Poisons

# Asbestos Fiber Amosite



***NYC FIREFIGHTER EXPOSED TO WTC DUST***

***ROGLI ET AL., AM J RESP CRIT CARE MED 166:797-800, 2002***

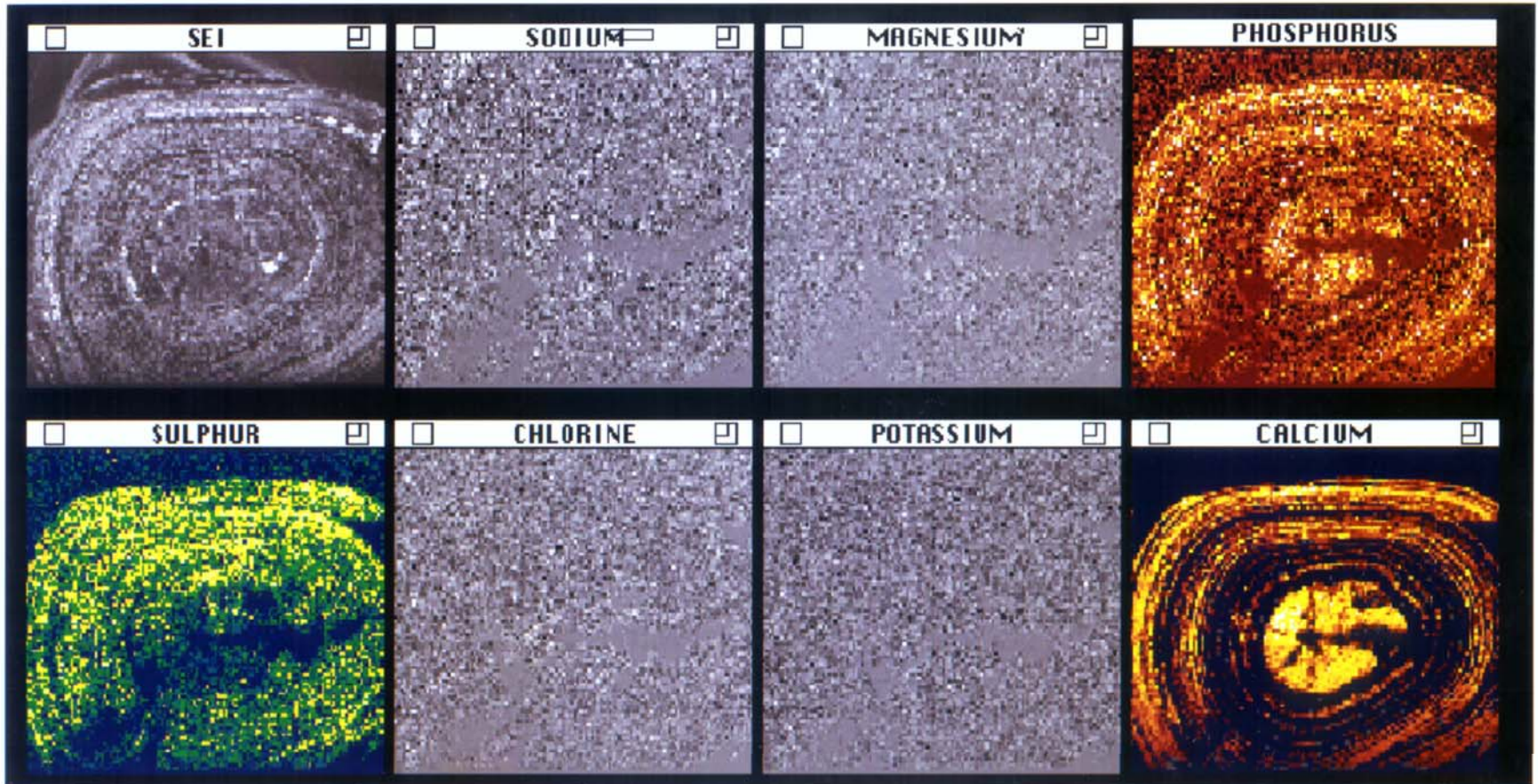
# Microanalysis as a Diagnostic Standard of Care

- Renal Stone Disease
  - Calcium Oxalate
  - Calcium Phosphate
  - Magnesium Ammonium Phosphate
  - Uric Acid
  - Cysteine
  - Bilirubin

# RENAL STONE



# RENALSTONE





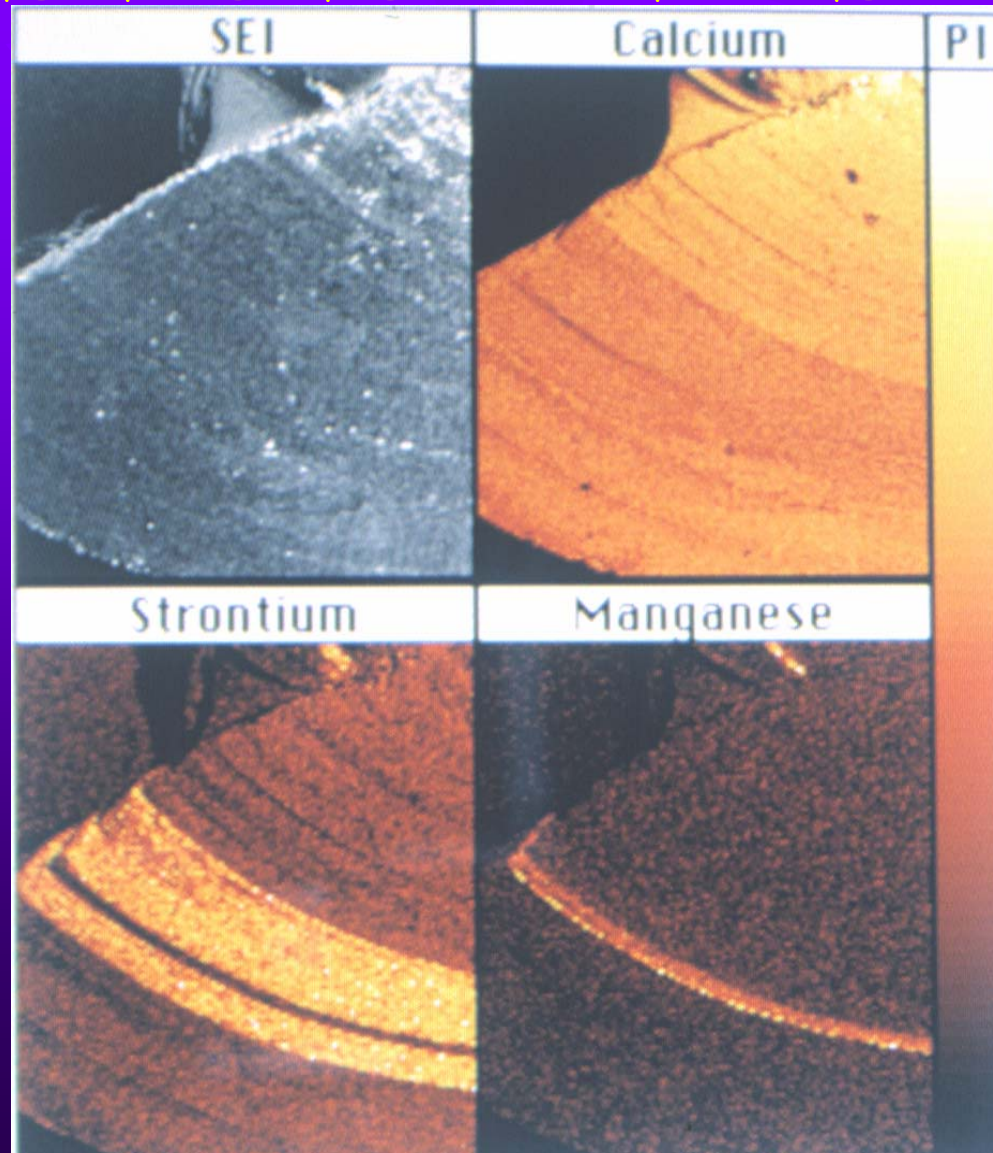
# Barnacle Biomineralization as Bioindicator

- Integrates water content (quality) over time
- Indicative of exposures to other organisms
- ❖ **Reveals time course and location of biomineralization front**

# Adult Barnacle Biomineralized Shell



# BIOMINERALIZATION IN THE PRESENCE OF METALS



**HIGH SPATIAL  
RESOLUTION  
GENERALLY REQUIRES  
THIN SECTIONS IN  
EPXMA OF  
BIOLOGICAL  
MATERIALS**



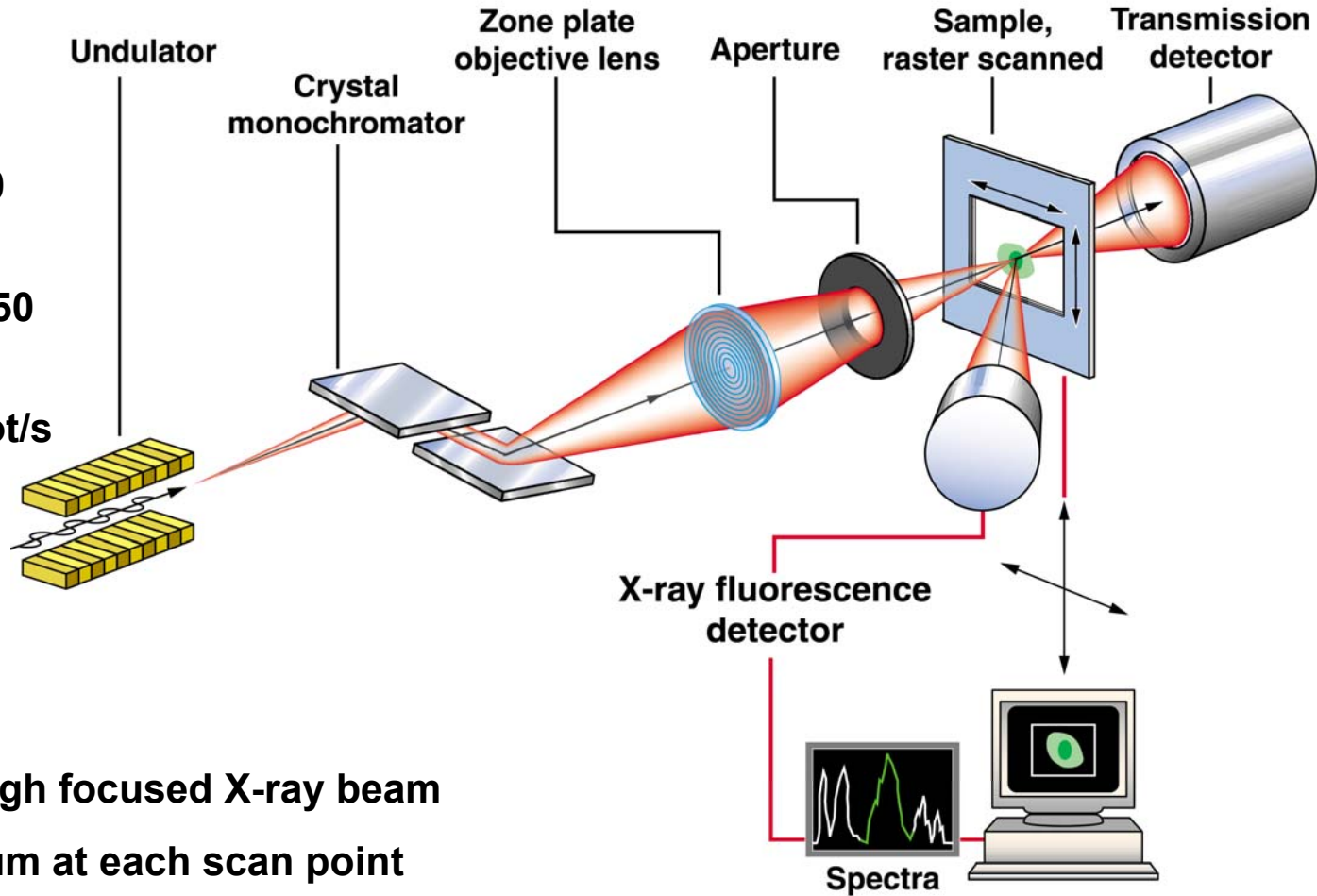
# XPXMA schematic

## Example APS 2-ID-D:

- energy range  $E = 5 - 30$  keV

- spatial resolution  $\delta = 150$  nm

- focussed flux  $2 \cdot 10^9$  phot/s  
(can trade flux for resolution)



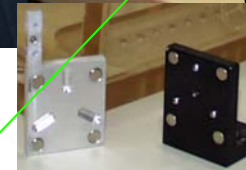
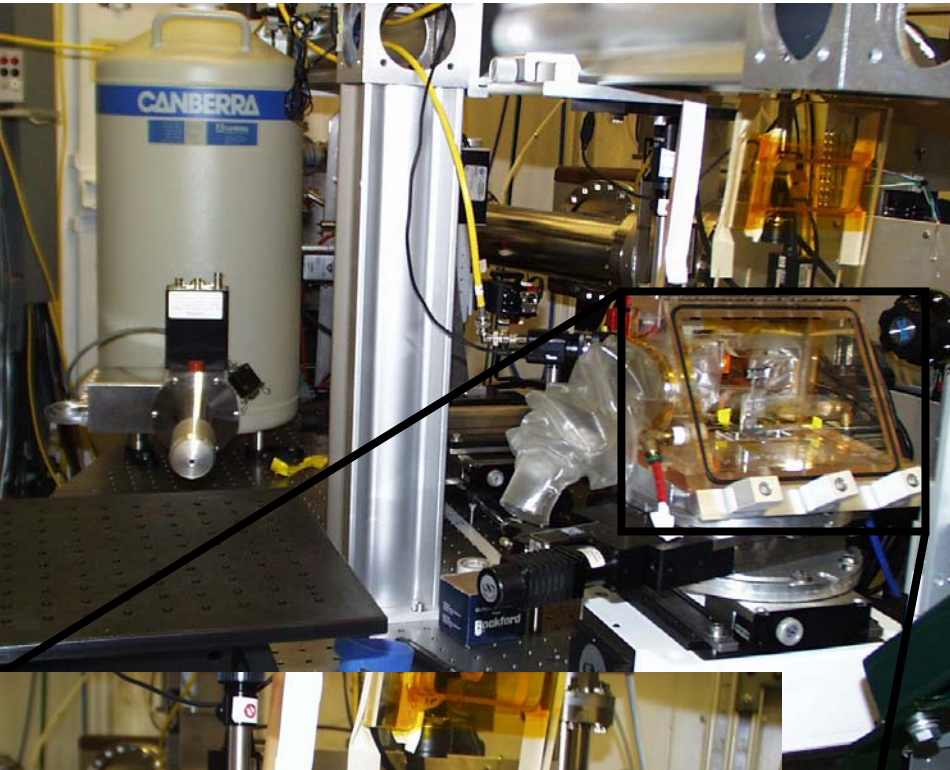
- stepscan sample through focused X-ray beam

- record full XRF spectrum at each scan point

- compare specimen counts/spectra to calibration curve, to quantify to area density

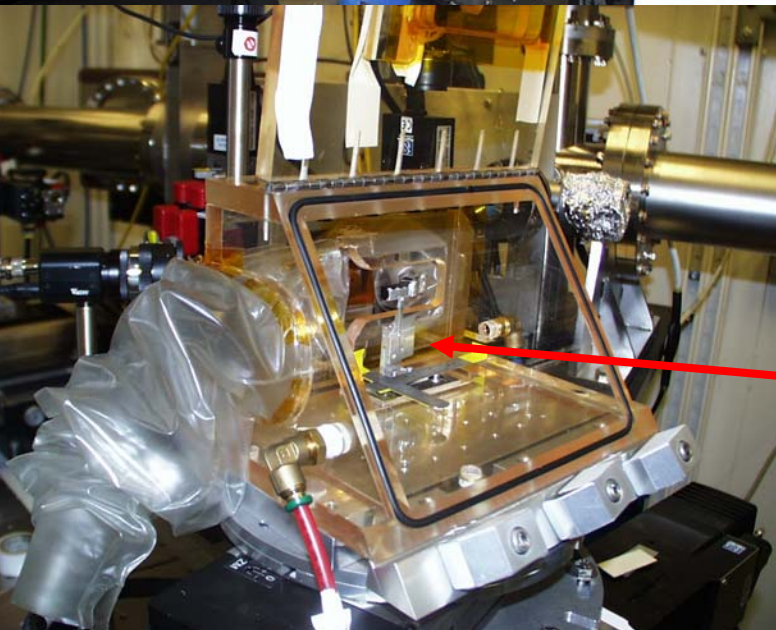
# 2-ID-E Hard X-ray Microprobe Facility

## Epi-Fluorescence Microscope

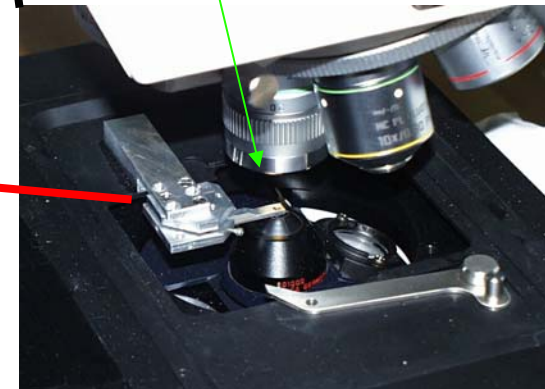


Sample Holder

Kinematic Mount



sample



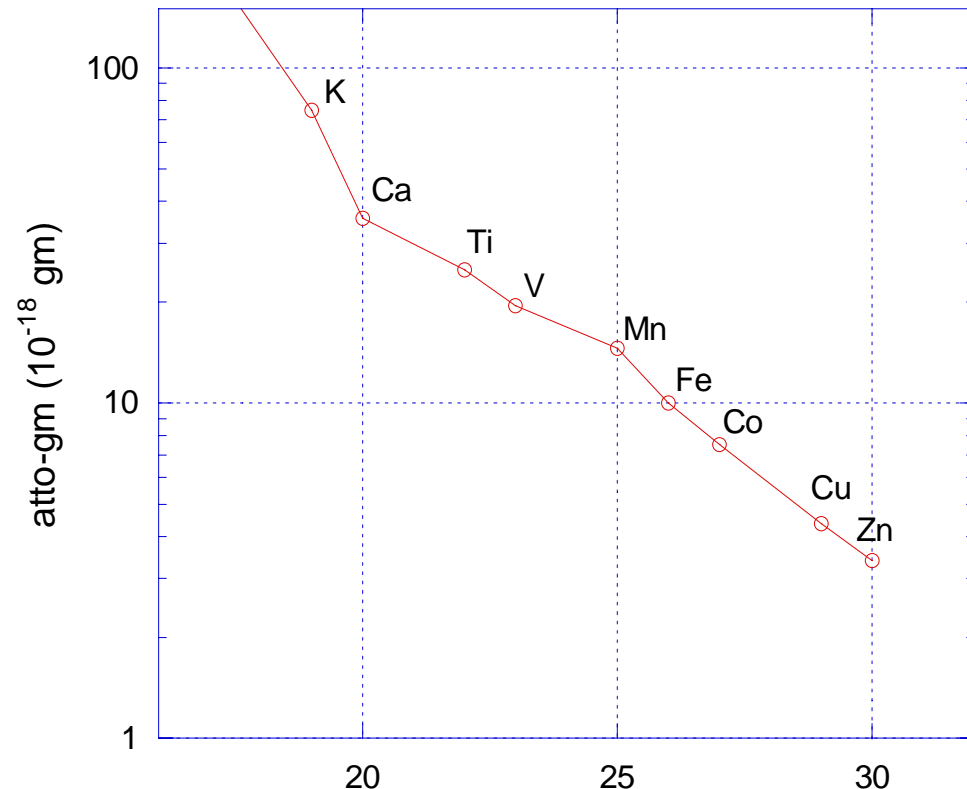
# **Trace elements / metals in biology & life sciences**

- essential cofactors in proteins**
- linked to diseases**
- in therapeutic drugs**
- as intracellular labels**
- contaminants in the environment with adverse impact on human health**



# Why use x-ray-induced fluorescence to study trace metals?

- Simultaneously map 15+ elements
- No dyes necessary
- very high sensitivity (sub part-per-million)
- quantitative
- large penetration depth (> 100  $\mu\text{m}$ )
- chemical state mapping & micro-XANES



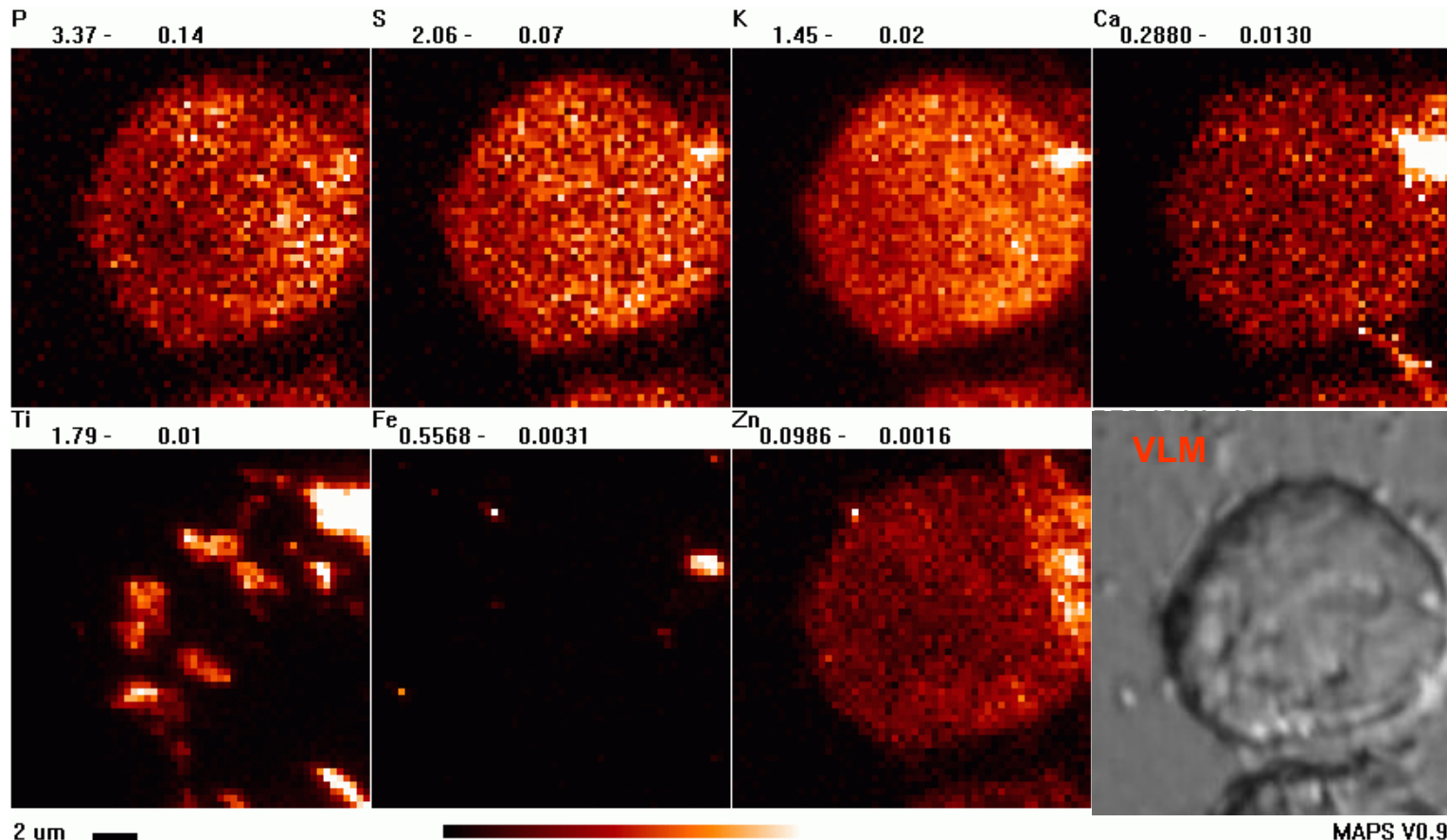
**Detection Limit for  
Transition Elements:  
for 1 sec. acquisition  
time, 0.2 x 0.2  $\mu\text{m}^2$  spot,  
E=10 keV**

# Example: Ti nanocomposites as intracellular probes

*T. Paunesku, G. Woloschak et al*

- attach inorganic  $\text{TiO}_2$  nanoparticle (4.5 nm diameter) to DNA strand to combine DNA specific biochemistry with semiconductor properties of  $\text{TiO}_2$
- carrier-particle that can bind to a specific chromosomal region w/ ability to release bound DNA & cause targeted strand breaks upon illumination

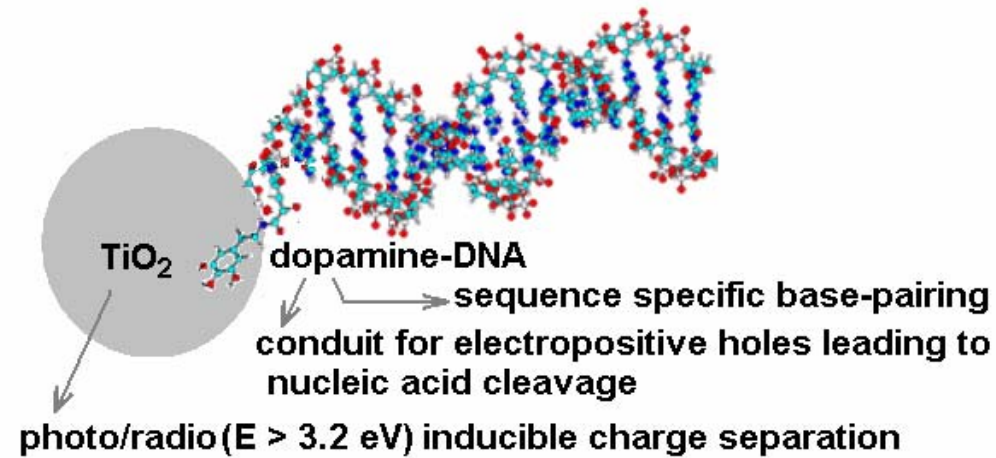
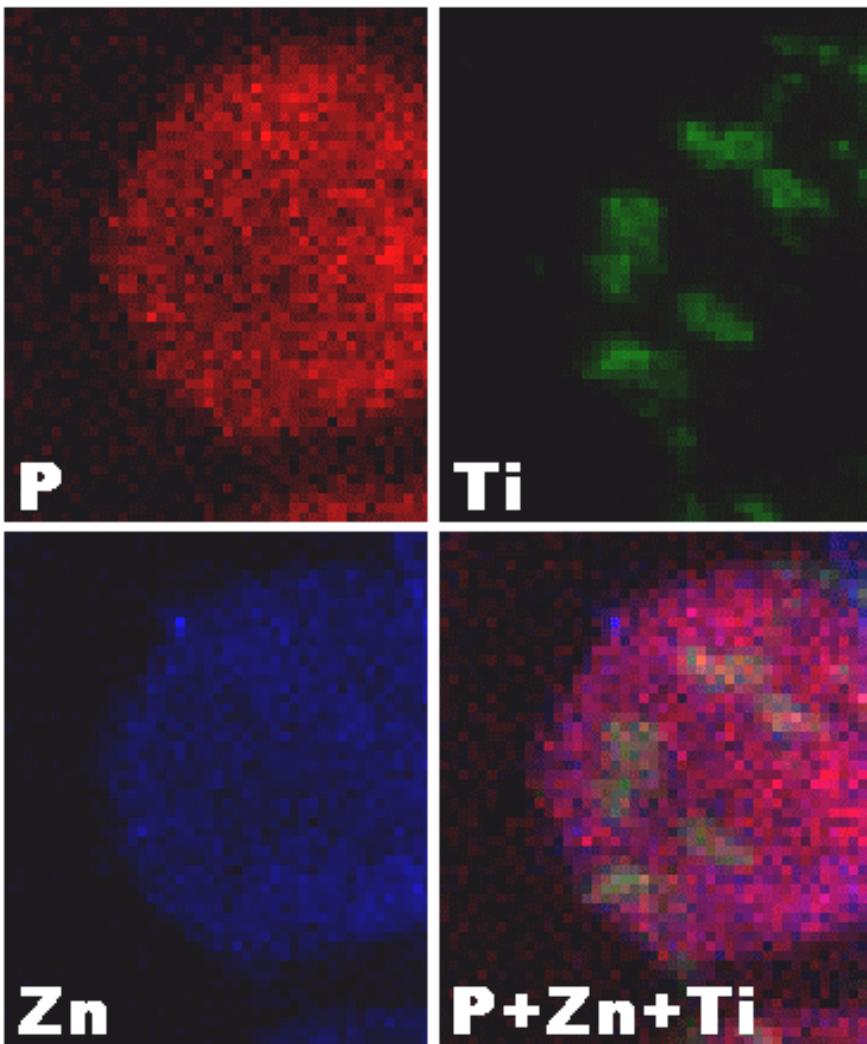
PC-12 cell  
transfected with  
nanocomposite  
combining  
mitochondrial  
DNA w.  $\text{TiO}_2$ .



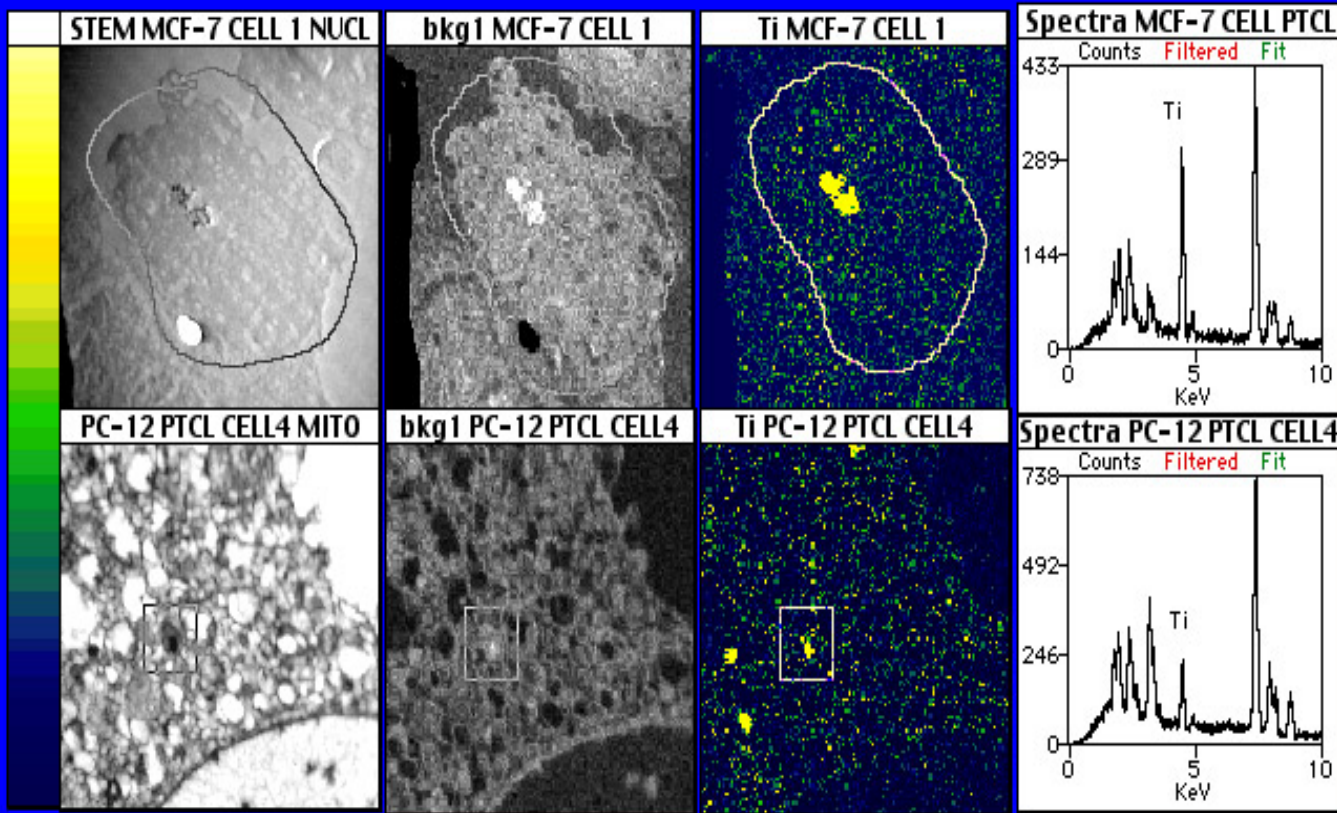
See also, T.  
Paunesku et al,  
*Nature Materials* 2,  
343-346 (01. May  
2003)

## Why study trace metals in life sciences:

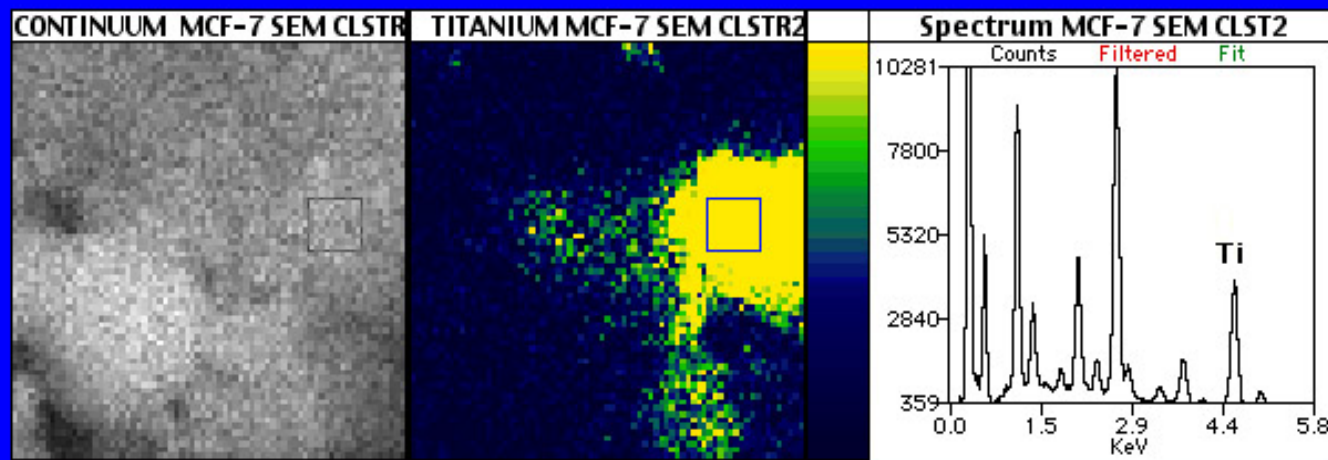
### Nanocomposites as intracellular tools



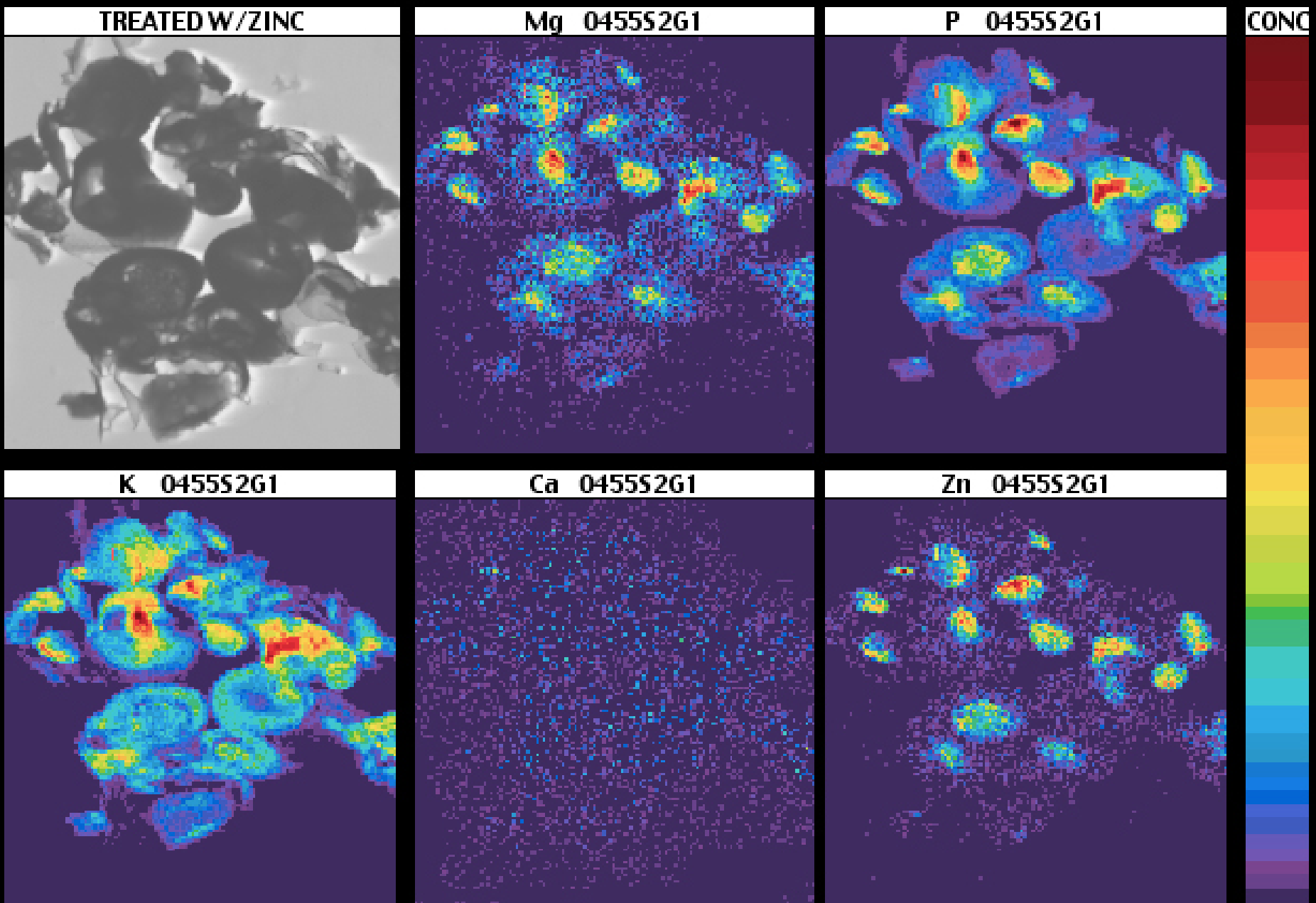
- attach  $\text{TiO}_2$  nanoparticle (4.5 nm diameter) to DNA
- combine DNA biochemistry with semiconductor properties of  $\text{TiO}_2$
- → carrier-particle that can bind to a specific chromosomal region w/ ability to cleave it upon illumination



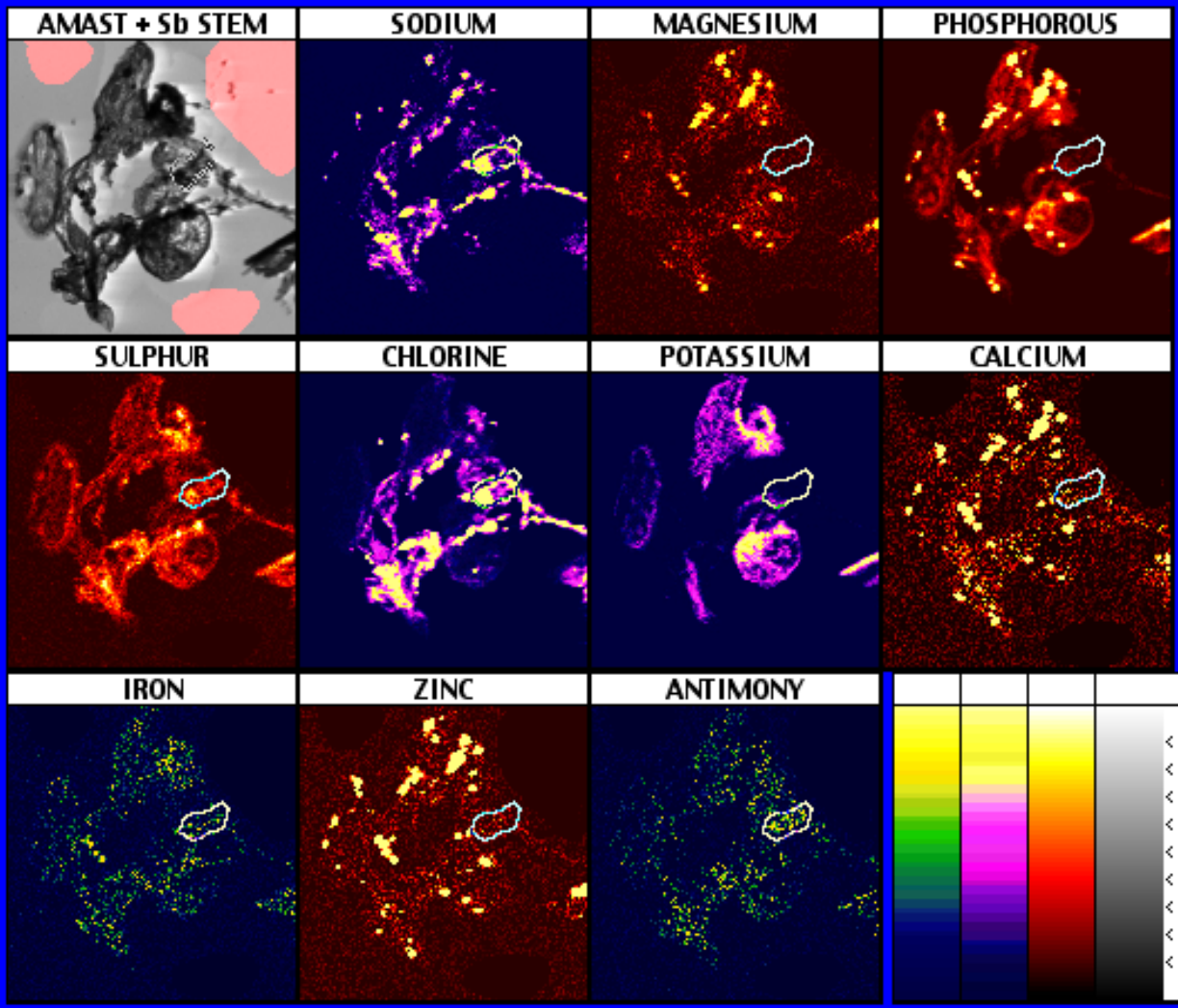
Elemental images and EPXMA spectra of MCF-7 breast cancer cells transfected with TiO<sub>2</sub> nanoparticles conjugated to **nuclear** DNA (top 4 panels) and to **mitochondrial** DNA (lower 4 panels). STEM at 80 KeV.



MCF-7 cells, similarly treated to those above, but air dried on to a carbon support and imaged with EPXMA in a scanning electron microscope at 20 KeV..

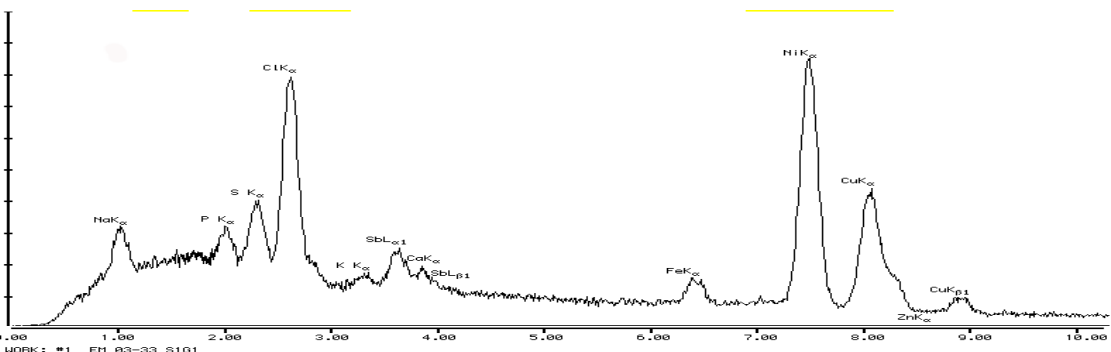


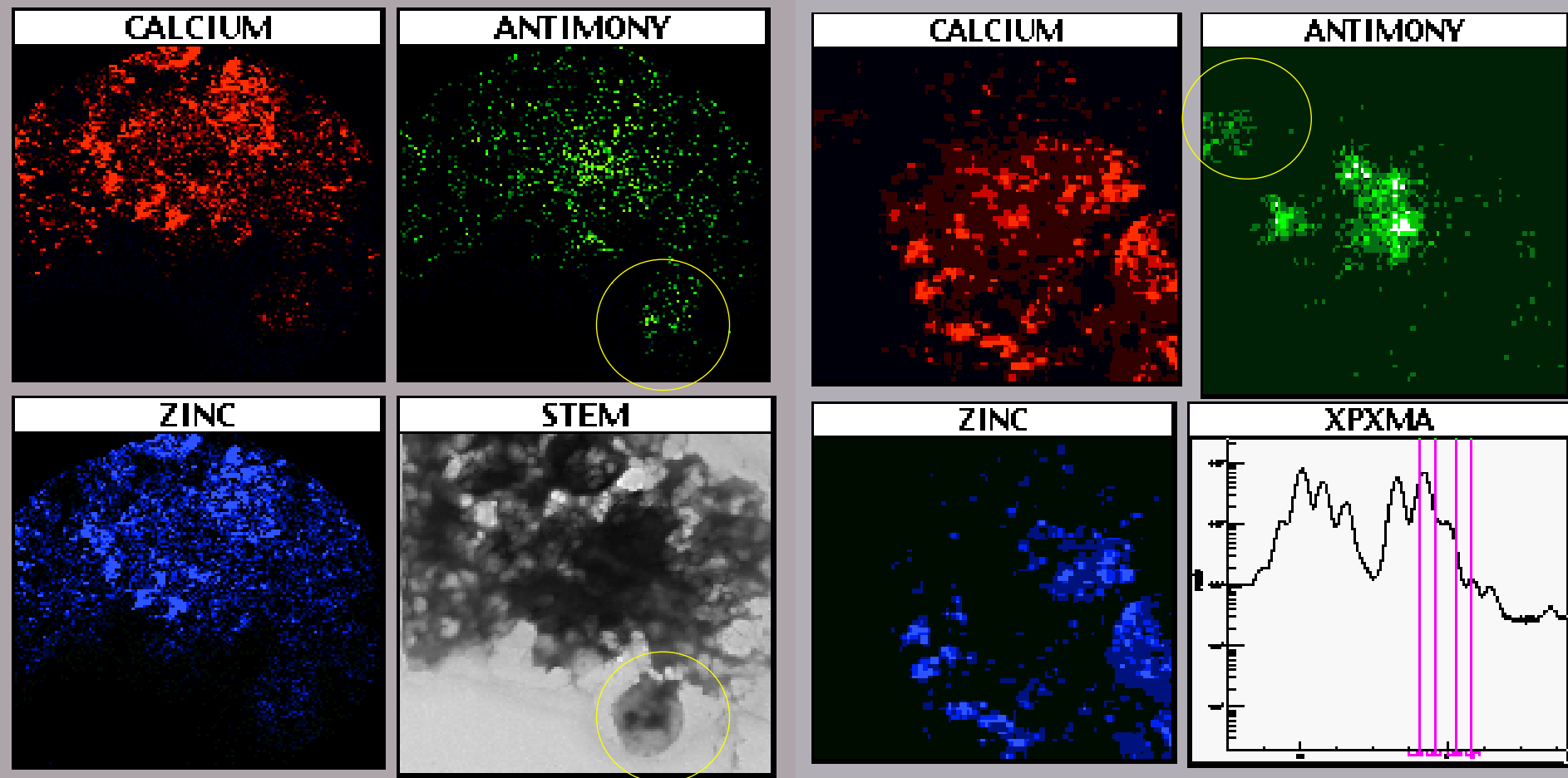
**EPXMA OF YEAST CELLS INCUBATED IN ZINC-RICH MEDIA**



Elemental EPXMA images from several cryoprocessed Sb-treated protozoa (*L. donovani*, amastigote stage).

The color scales are fully quantitative but have been adjusted to accentuate related elements. The spectrum was taken from the region outlined.





## EPXMA

## XPXMA

Comparative images of the same cells from several Sb-treated protozoa (*L. donovani*, amastigote stage 1000 $\mu$ g/ml SbV, for 1 hr).. The intensity of the colors qualitatively reflect the content of the elements. The yellow circled region is from the same area of the specimen. Note that the EPXMA set on the left is a mirror image of the XPXMA set on the right.

MICROANALYSIS METHOD	ELECTRON MICRO-PROBE	PROTON MICRO-PROBE	SIMS MICRO-PROBE	LAMMA MICRO-PROBE	X-RAY MICRO-PROBES
MDL (ppm)*	100	0.5	0.005	0.005	0.01
MDL ( $\mu$ M)	75	3	0.003	0.003	0.05
QUANTITATION	GOOD	GOOD	V. DIFFICULT	POSSIBLE	GOOD
RESOLUTION ( $\mu$ m)	0.01	1.0	0.04	0.5	0.03** 0.05#
HYDRATED SAMPLES	NO##	DIFFICULT	NO	DIFFICULT	POSSIBLE
BEAM DAMAGE	HIGH	MED	HIGH	HIGH	LOW

\*In part, after Spanne and Rivers (1987)

\*\*Predicted for the APS Bio-Nanoprobe

##Except for water content from frozen specimens

# Transmission Imaging has demonstrated  $\sim 0.05 \mu\text{m}$  between 6 to 8 keV. Bilderback et al. (1994)

## Comparison of XPXMA with other sub-cellular biological microprobe analysis methods